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(54) Title: SOLUTION FOR TREATING AUTOLOGOUS TISSUE FOR IMPLANTATION

(57) Abstract: An aqueous solution containing a water miscible organic solvent, polyethylene glycol and heparine, is used to modify the tissue reactivity of an autologous tissue freshly obtained from a host mammal and to render the tissue temporarily more rigid than in its native state, and better suited for shaping, moulding, handling and cutting prior to implantation into the host patient. The implant is highly resistant or immune to thickening, contraction and reduced fibrin deposition after it is implanted and exposed to the bloodstream of the host.

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SOLUTION AND METHOD FOR TREATING AUTOLOGOUS TISSUE FOR IMPLANT OPERATION BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention is in the field of methodology and materials for performing autologous tissue transplants. More particularly, the present invention is directed to a solution suitable for treating autologous tissue after the tissue has been removed from a mammal, to render the tissue more suitable for handling and molding it into the desired shape and obtain predictable results after implantation in the mammal.

2. Brief Description of the Prior Art

Significant advances have been made in the field of treatment of defective heart valves due to abnormalities during fetal development, or due to infectious or degenerative diseases. These surgical treatments most often require the use of biocompatible materials that can be either synthetic polymers or of biological origin, either from the patient (autologous), an individual of the same species (homologous), or different species (heterologous or xenograft). Defective heart valves are replaced with mechanical valves or tissue valves, such as cadaver or animal aortic valves (bioprosthesis). Because of their more-or-less predictable mechanical wear properties, the mechanical prostheses have been proven suitable for their intended purpose, primarily in younger patients. However, mechanical prostheses have their disadvantages because patients having them require long-term, constant and vigorous anti-coagulant therapy. In older patients however the bioprosthesis have been favored mainly because they do not require anti-coagulation therapy, and in older patients they do not tend to undergo calcification as often as they tend to do in younger patients. Cryo-preserved homografts have been used widely in the western world during the recent years. However, these are hard and expensive to obtain, ship and store,

1 and their availability on a world-wide basis appears to be limited.

2 As is known, heart valve repair or replacement and many other implant
3 operations require soft connective tissue which in preparation for implantation
4 needs to be sized and cut into specific shapes. A substitute for such soft
5 connective tissue of biological origin can be provided by flat sheets of certain
6 synthetic materials. However, it is difficult to find synthetic materials which
7 can match the compliance of the native tissue they are intended to replace, and
8 which do not engender adverse reaction by the recipient of the implant.
9 Autologous tissues, such as pericardium, hold the promise for an ideal soft
10 tissue replacement material in implants, and fresh autologous pericardium has
11 been used in the prior art as a tissue source for repairing a variety of heart
12 lesions, including heart valves. However, results with the use of such fresh
13 untreated autologous tissue were less than satisfactory because tissue
14 contraction distorted the repair, and in case of heart valves, tended to render
15 the leaflets non-functional some time after operation. Generally speaking, the
16 problem with fresh autologous soft connective tissue, such as pericardium, is
17 that such tissues are often too soft and flexible to cut and otherwise handle
18 especially during open heart surgery where an atmosphere of urgency prevails.
19 As an improvement Dr. Duran (one of the inventors of the present invention)
20 developed a procedure in which the autologous tissue that has been freshly
21 obtained from the patient operated on, is treated with 0.5% glutaraldehyde in a
22 mold for 10 minutes. Thereafter, it is cut into the desired shape dictated by
23 the mold and is placed in the patient as new replacement heart valve leaflets.
24 Although this procedure works reasonably well, the disadvantage of tissues
25 treated by glutaraldehyde is that, similarly to xenografts, such tissues may
26 well undergo calcification in long term implants.

27 The present invention provides an alternative to glutaraldehyde fixation
28 of autologous tissues and yet eliminates the problems caused by contraction of
29 fresh tissue and the difficulty of handling and manipulating soft tissue.

1 Because the invention avoids the above-noted problems by treating the
2 autologous tissue with an aqueous solution of alcohols and other materials,
3 prior art describing solutions and methods for treating biological tissues and
4 specimens are thought to be of interest as background to the present invention.
5 Such prior art can be found in United States Patent Nos. 5,558,875;
6 5,296,514; 5,276,006; 4,323,358 and 4,329,492. Among the foregoing, the
7 most recently issued United States Patent No. 5,558,875 describes a process
8 of preparing a collagenous prosthesis by soaking tissue in an organic detergent
9 for sufficient time to disrupt the cell membrane and to solubilize the cellular
10 membrane proteins of the collagenous tissue and thereafter extracting and
11 removing the cellular membrane proteins from the collagenous tissue by
12 mechanical washing to obtain the prosthesis and thereafter preserving the
13 prosthesis in alcohol. The process is said to preserve the elasticity of the
14 prosthesis.

15 The following articles or scientific publications also provide
16 background of interest to the present invention: *Chachques et al.*, Ann. NY
17 Acad Sci. 1988, 529:184; *Love et al.*, J. Heart Valve Dis 1992: 1:232-41;
18 *Chauvaud et al.*, J. Thorac Cardiovasc Surg. 1991, 102:171-8; *Duran et al.*, J.
19 Thorac Cardiovasc Surg. 1995, 11-511-6; *Vyavahare et al.*, 4th Scientific
20 Meeting International Association for Cardiac Biological Implants,
21 Washington DC, May, 1997; *Ritter et al.*, Plastic & Reconstructive Surgery.
22 101(1):142-6, Jan., 1998; and *Vetter et al.*, J. Thorac Cardiovasc Surg,
23 35(1):11-5, Feb. 1987.

24 SUMMARY OF THE INVENTION

25 It is an object of the present invention to provide a composition
26 (solution) and method for treating autologous soft tissue so as to render it
27 easier to handle and shape for implantation, while avoiding disadvantages
28 caused by aldehyde treatment of such tissue.

29 It is another object of the present invention to provide a composition

1 (solution) and method that meets the foregoing objective and which treats the
2 autologous soft tissue during a surgical procedure and while said procedure is
3 in progress.

4 The foregoing and other objects and advantages are attained by
5 exposing for approximately 2 to 8 minutes a fresh autologous tissue, such as
6 pericardium, to an aqueous solution containing approximately 10 to 70 % by
7 volume of a water-miscible non toxic polar solvent, such as ethyl alcohol,
8 approximately 2 to 30 % by weight of polyethylene glycol of a molecular
9 weight between approximately 6,000 to 15,000 D, and approximately 0.01 to
10 1.0 % by weight of heparin. The tissue preferably, and most frequently in
11 accordance with the procedure is immersed in the above-described solution
12 while placed in a suitable mold. In case of preparing the tissue for heart valve
13 replacement the mold is configured to provide the appropriate shape and
14 dimension for the replacement heart valve leaflets. The soft tissue implant
15 treated in the foregoing manner temporarily becomes more rigid and easier to
16 handle during surgical procedure than unprepared fresh tissue. However,
17 within approximately the time taken to perform the surgical procedure of
18 implantation the treated tissue regains its original physical properties,
19 including its elasticity.

20 BRIEF DESCRIPTION OF THE DRAWING FIGURES

21 **Figure 1** is a schematic perspective view showing the general
22 configuration of one cusp of a negative template of a mold used for shaping
23 an aortic valve replacement from autologous tissue, utilizing the novel solution
24 and method of the present invention.

25 **Figure 2** is a schematic perspective view showing the general
26 configuration of one cusp of a positive template of the mold used for shaping
27 an aortic valve replacement from autologous tissue, utilizing the novel solution
28 and method of the present invention.

29 **Figure 3** is a top plan view of the negative cusp of **Figure 1**.

1 **Figure 4** is an end plan view of the negative cusp of **Figure 1**.

2 **Figure 5** is a side plan view of the negative cusp of **Figure 1**.

3 **Figure 6** is a schematic top plan view showing the general
4 configuration of three negative cusps of **Figure 1** assembled to form a
5 negative template used for shaping an aortic valve replacement from
6 autologous tissue, utilizing the novel solution and method of the present
7 invention.

8 **Figure 7** is a front plan view of the negative template of **Figure 6**.

9 **Figure 8** is a schematic perspective view of the negative template of
10 **Figure 6**.

11 **Figure 9** is a detailed top plan view of a first preferred embodiment of a
12 negative template used for shaping a pericardial valve replacement from
13 autologous tissue, utilizing the novel solution and method of the present
14 invention.

15 **Figure 10** is an end view of the first preferred embodiment of the
16 negative template of **Figure 9**.

17 **Figure 11** is a front plan view of the first preferred embodiment of the
18 negative template of **Figure 9**, with part of the front material broken away.

19 **Figure 12** is a detailed top plan view of a first preferred embodiment of
20 a positive template used for shaping a pericardial valve replacement from
21 autologous tissue, utilizing the novel solution and method of the present
22 invention.

23 **Figure 13** is an end view of the first preferred embodiment of the
24 positive template of **Figure 12**.

25 **Figure 14** is a front view of the first preferred embodiment of the
26 positive template of **Figure 12**.

27 **Figure 15** is a perspective view of the first preferred embodiment of the
28 negative template of **Figure 9**.

29 **Figure 16** is a perspective view of the first preferred embodiment of the

1 positive template of **Figure 12**.

2 **Figure 17** is a top plan view showing the first preferred embodiment of
3 the negative template of **Figure 9** and the first preferred embodiment of the
4 positive template of **Figure 12** assembled to one another.

5 **Figure 18** is a front plan view showing the first preferred embodiment
6 of the positive template of **Figure 9** and the first preferred embodiment of the
7 negative template of **Figure 12** assembled to one another.

8 **Figure 19** is a partial cross-sectional view taken on lines 19,19 of
9 **Figure 18**.

10 **Figure 20** is a view showing the assembled mold of **Figure 18** having
11 autologous tissue and immersed in a solution in accordance with the present
12 invention.

13 **Figure 21** is a partial cross sectional view of the mold with autologous
14 tissue, the cross-section being taken on lines 21,21 of **Figure 20**.

15 **Figure 22** is a partial schematic view, schematically showing the
16 trimming of excess autologous tissue to form a replacement heart valve, in
17 accordance with the present invention.

18 **Figure 23** is a cross-sectional view taken on lines 23,23 of **Figure 22**.

19 **Figure 24** is a partial cross sectional view of the trimmed autologous
20 tissue.

21 DESCRIPTION OF THE PREFERRED EMBODIMENTS

22 The present invention is practiced in conjunction with a surgical
23 procedure wherein a heart valve, aortic, pulmonary, tricuspid or mitral, or
24 other biological membrane is replaced or repaired. Because its most frequent
25 use is in conjunction with replacement of defective membranes in the heart,
26 the present invention is described here primarily as it pertains to replacement
27 of defective heart valves. In accordance with the present invention the
28 operating surgeon excises a membrane-like fresh autologous tissue from the
29 patient and by application of the solution of the present invention changes the

1 physical properties of the fresh autologous tissue to be better suited for
2 trimming, handling, and manipulation during implantation into the patient.
3 Membrane-like tissues which are suitable to be handled and implanted in
4 accordance with the present invention include the peritoneum, pericardium,
5 gut, dermis pleura and tendon. For open heart surgery and replacement of
6 defective heart valves the use of the patient's pericardium is preferred, and
7 therefore the invention is described herein primarily in connection with the use
8 of pericardium as the membrane-like autologous tissue.

9 In accordance with the invention, the freshly obtained pericardium (or
10 other membrane-like autologous tissue) is treated with an aqueous solution
11 containing approximately 10 to 70 % by volume of a water-miscible non-toxic
12 organic solvent, approximately 2 to 30 % by weight of polyethylene glycol of
13 a molecular weight between approximately 6,000 to 15,000 D, and
14 approximately 0.01 to 1.0 % by weight of heparin, the rest of the solution
15 being water. Examples of suitable water-miscible organic solvents or liquids
16 are lower alkyl, especially C₁ to C₃ alcohols, such as methanol, ethanol and
17 *iso*-propanol, and acetone, acetonitrile and methyl ethyl ketone. A more
18 preferred range of the components in the solution in accordance with the
19 present invention is approximately 15 to 60 % by volume of the water-
20 miscible organic liquid, 2 to 10 % by weight of polyethylene glycol, and 0.1 to
21 0.7 % by weight of heparin.

22 Preferably the organic solvent is ethyl alcohol, and in the presently
23 most preferred embodiment of the solution there is approximately 50 % by
24 volume ethanol, approximately 5 % by weight of polyethylene glycol having a
25 molecular weight of approximately 8,000 D, and approximately 0.5 % by
26 weight of heparin. The biological membrane is thoroughly exposed to the
27 solution for sufficient time to provide the desired results of rendering the
28 membrane more rigid and therefore easier to trim, suture and otherwise
29 handle. However, usually a time limit is set to this exposure by the fact that

1 the process occurs while the patient is undergoing surgery, usually open heart
2 surgery. It was found in practice that approximately 2 to 8 minutes of
3 exposure of the biological membrane to the solution is sufficient.
4 Nevertheless, under circumstances where the surgical procedure *per se* does
5 not represent a time-limiting factor, the biological membrane can be kept in
6 the solution for indefinite length of time provided the solution is kept under
7 sterile condition. Treatment by this solution kills the living cells in the
8 membrane although treatment with the solution containing organic solvent at
9 the lower end of the above-described range may only kill cells on the surface
10 of the membrane and merely retards the biological response of cells in the
11 interior. Nevertheless, unlike treatment with glutaraldehyde, treatment with
12 the solution of the present invention does not result in any cross-linking of the
13 membrane materials. The biological membrane or tissue becomes more rigid
14 or stiff during exposure to the solution partly because of the hypertonic,
15 dehydrating nature of the solution.

16 Hardening or stiffening of the membranes is temporary, however,
17 because after sufficient rinsing with saline or upon equilibration with isotonic
18 biological fluids, such as blood, the biological membranes regain virtually
19 completely their original physical properties, and as a result are well suited for
20 their intended function as replacement of natural membranes, primarily as
21 heart valves.

22 A preferred manner of practicing the present invention, together with
23 molds that are used for shaping pericardium or other biological membranes to
24 provide aortic and pericardial heart valve replacements are illustrated in the
25 drawing figures. Referring now back to the Brief Description of the Drawing
26 Figures, **Figures 1 through 8** schematically illustrate the basic geometry of a
27 mold comprising a negative **30** and a positive **32** template for forming an
28 aortic valve replacement in accordance with the present invention. **Figures 1**
29 **through 5** schematically illustrate the basic geometry of the individual negative

1 34 and positive 36 cusps of the templates 30 and 32 that together form the
2 mold. The templates 30 and 32 are made from thin plastic material, and are
3 configured and dimensioned to provide the aortic heart valve replacement for
4 the individual patient who is being operated on. As it will be readily
5 understood by those skilled in the art of cardiology and related cardiac
6 surgery, primarily echocardiograms of the patient provide the information as
7 to what size heart valve replacement is needed. Edges of the templates 30 and
8 32 are beveled or rounded in order to facilitate trimming of excess tissue with
9 a surgical knife.

10 In accordance with one manner of practicing the invention, the
11 pericardium (or other suitable biological membrane) is placed into the mold
12 between the negative and positive templates and treated for approximately 5
13 minutes with the solution of the invention by immersion in the solution.
14 Thereafter it is very quickly (for less than 5 seconds) rinsed with saline
15 solution containing approximately 250 unit per ml heparin. This step of
16 treating the tissue with heparin solution for a very brief period of time is not
17 necessary for the successful practice of the invention and is therefore optional.
18 In any event, after removal from the mold and having been treated with the
19 solution of the invention the tissue is more rigid than the native untreated
20 pericardium, and is easier to handle. Excess tissue is then removed by
21 trimming with a surgical knife, the tissue in the shape of heart valve leaflets is
22 removed from the mold, and is thereafter surgically implanted. The
23 increased rigidity or stiffness of these replacement leaflets renders the
24 implantation procedure easier to handle. After suturing is completed, the
25 tissue is irrigated with saline solution, whereupon it regains its original
26 physical properties. As noted above, the biological membrane treated in
27 accordance with the present invention regains its original physical properties
28 upon adequate rinsing with saline, or achieving equilibrium with isotonic
29 aqueous fluid, such as blood.

1 **Figures 9 through 24** illustrate in more detail an actual mold
2 comprising a negative template **38** and a positive template **40** adapted for
3 shaping a flat biological membrane, such as pericardium, to form replacement
4 leaflets for a pericardial valve. The two templates **38** and **40** of this mold are
5 made of thin plastic material having beveled edges, and the templates are
6 dimensioned to provide replacement leaflets of appropriate size for the patient
7 who is undergoing the open heart surgery. For use in conjunction with the
8 present invention, the negative **38** and the positive template **40** both are
9 provided with a plurality of apertures **42**. When this mold is used in the
10 practice of the present invention, the autologous biological membrane,
11 preferably pericardium excised from the patient who is undergoing open heart
12 surgery, is placed between the templates, **38** and **40**, that is into the mold.
13 Accordingly, the pericardium, shown in **Figures 21 through 24** as **44** is
14 sandwiched between the two templates **38** and **40**. The two templates of the
15 mold are held together with suitable plastic clips **46**, shown in **Figure 20**.
16 The mold including the pericardium **44** is then immersed for approximately 5
17 minutes in the solution **48** of the invention, as is schematically shown in
18 **Figure 20**. During this time the solution **48** percolates through the apertures
19 **42** into the pericardium **44** and renders the pericardium **44** more rigid than in
20 its natural native state. After removal from the solution, the tissue **44** is
21 trimmed with a surgical instrument along the beveled edges of the mold. The
22 surgical instrument is schematically shown in **Figure 22** and bears the
23 reference numeral **50**. The resulting replacement heart leaflets (not shown) are
24 then removed from the mold, and may be quickly (less than 5 seconds) rinsed
25 with saline solution containing approximately 250 unit per ml heparin. This
26 optional quick rinsing does not yet decrease the rigidity of the tissue which
27 was the result of treatment with the solution. Surgical implantation of the
28 replacement leaflets is greatly facilitated by its increased rigidity or stiffness.
29 After implantation, the replacement leaflets are irrigated with saline and regain

1 their original physical properties. A substantial advantage of the heart valve
2 replacements obtained in accordance with the present invention is that, unlike
3 replacement valves made of autologous tissues in the prior art, the replacement
4 valves of the invention do not contract or shrink after implantation. Generally
5 speaking, implants of autologous tissues which have been treated in
6 accordance with the present invention are highly resistant or immune to
7 thickening, contraction or fibrin deposition after the implants are placed into
8 the bloodstream of the host. The above described process of exposing the
9 biological membrane to the solution of the invention while the membrane is
10 held in an appropriately configured and dimensioned mold is the presently
11 preferred mode of practicing the invention.

12 Specific Examples

13 Fresh autologous tissues were dissected from the sheep and placed on
14 molds which were used as templates for cutting the tissues into a shape which
15 appeared as three half-moons joined together at the base of the half-moon.
16 This particular shape was designed for the purpose of aortic or pulmonary
17 heart valve cusp extension operation. Pericardial tissues cut according to this
18 design can be directly implanted into the heart of the individual patient where
19 the tissue is derived from. The tissues in the mold were immersed into a
20 solution containing 50 % by volume of alcohol, 5 % by weight of polyethylene
21 glycol (MW=8,000) and 0.5 % by weight of heparin for five minutes at room
22 temperature. The tissues in the mold were removed from the solution and
23 after the removal of excess liquid outside the tissues, the tissues were rinsed in
24 saline containing 250 unit/ml of heparin for less than 5 seconds.

25 Tissues treated with the solution mentioned above appeared to be
26 slightly translucent and natural in color. Unlike the fresh untreated tissues, the
27 treated tissues were stiffer so that it was relatively easy to lift the tissues
28 without the tissues folding onto themselves. The treated tissues were easily
29 spread on a flat or curved surface with different markings for cutting the

1 tissues. Since the cut tissues maintained their shape, they were easily
2 implanted as replacement leaflets of heart valves. Yet, once the suturing of the
3 tissues was completed and a small amount of saline was irrigated onto the
4 implanted tissues, the tissues became soft quickly. Within the time required to
5 close the open heart, the physical properties of the tissues became
6 indistinguishable from the fresh untreated tissues. Therefore the resulting
7 implants function perfectly as repaired heart valves.

8 Human fibroblasts and umbilical cord vein endothelial cells were
9 cultured on the treated tissues after there were rinsed in saline containing 250
10 unit/ml of heparin to study their biocompatibility. Round discs of the tissues
11 were cut to fit the bottom of the wells of a 24 well culture plate. Tygon^R
12 flexible rings were placed on top of the tissues to ensure a good seal at the
13 edge of the tissues. Cells were seeded on the tissues in normal culture media
14 for one week. At the end of the incubation period, tissues were recovered and
15 processed for histology. Both human umbilical cord vein endothelial cells and
16 human skin fibroblasts attached and proliferated on the treated tissues as
17 evidence that after rinsing in saline the treated tissue is not cytotoxic and
18 biocompatible for host cells to attach and proliferate. The attachment and
19 proliferation of endothelial cells and other connective tissue cells on cardiac
20 implants is potentially important for the long term survival of the implant.

21 Integrity of the collagen fibers in the treated tissues was examined by
22 melting temperature measurements. Tissues were heated in phosphate
23 buffered saline from 37° C until they shrunk. The shrinkage temperature of
24 the treated tissues after they were rinsed in saline was 64±1 °C which is
25 identical to untreated fresh tissue indicating that the collagen fibers remained
26 intact throughout the treatment and saline-rinse process.

27 Efficacy of the treated tissues as useful cardiovascular implants was
28 tested by implanting the treated autologous tissues in the descending aorta of
29 sheep in different configurations. A piece of pericardium was dissected and

1 divided into three pieces with different shapes, namely a trapezoid, a strip
2 made into a conduit and a square. These pieces of tissues were implanted
3 serially in the descending aorta of sheep. The trapezoid shaped tissue was
4 implanted upstream as a patch on the aortic wall, next to it down-stream the
5 short conduit was implanted and further down-stream a square shaped tissue
6 was placed as a semi-free flap across the lumen inside the aorta with two edges
7 of the square attached to opposite side of the inner wall of the aorta. When
8 fresh autologous tissues were implanted under the same condition, the semi-
9 free flap in the aorta lumen became fibrotic and contracted within 30 days.
10 However the patch and the conduit upstream, that was implanted in
11 accordance with the present invention did not show the same reaction. There
12 was also evidence of thrombus formation and fibrin deposition on the surfaces
13 of the fresh implants. When the treated implants (not rinsed in saline) were
14 implanted in sheep in the same manner all implants remained intact after 30
15 days without any evidence of fibrotic reaction and tissue contraction.
16 Thrombus and fibrin deposition were minimal or absent on these implants.

17 In still other further examples fresh bovine pericardial tissues were
18 treated using the solution of the invention. The treated tissues were cut and
19 trimmed to sizes and shapes suitable for valve repairs. The treated and
20 trimmed tissues were sutured in the aortic roots of isolated human and porcine
21 hearts. The whole hearts were then mounted on a pulse duplicator to examine
22 the competency of the repair valve. The treated tissues were very flexible and
23 the reconstructed valves functioned normally as competent aortic valves.

WHAT IS CLAIMED IS:

- 1
2 1. A liquid composition adapted for treating autologous biological
3 tissue for modifying its tissue reactivity and for rendering the tissue
4 temporarily more rigid than in its natural state, the composition comprising:
5 approximately 10 to 70 % by volume of a water miscible non-toxic
6 organic solvent selected from the group consisting of an alcohol having 1 to 3
7 carbons, acetone, acetonitrile and methyl ethyl ketone;
8 approximately 2 to 30 % by weight of polyethylene glycol having a
9 molecular weight in the range of approximately 6,000 to 15,000 D;
10 approximately 0.01 to 1.0 % by weight of heparin, and
11 the balance of the composition substantially consisting of water.
12 2. The liquid composition of Claim 1 wherein the water miscible
13 organic solvent is ethyl alcohol.
14 3. The liquid composition of Claim 2 that contains approximately
15 15 to 60 % ethyl alcohol.
16 4. The liquid composition of Claim 3 that contains approximately
17 50 % ethyl alcohol.
18 5. The liquid composition of Claim 1 that contains approximately 2
19 to 10 % polyethylene glycol.
20 6. The liquid composition of Claim 5 that contains approximately 5
21 % polyethylene glycol.
22 7. The liquid composition of Claim 5 wherein the polyethylene
23 glycol has a molecular weight of approximately 8,000 D.
24 8. The liquid composition of Claim 1 that contains approximately
25 0.1 to 0.7 % heparin.
26 9. The liquid composition of Claim 8 that contains approximately
27 0.5 % heparin.
28 10. An aqueous liquid composition adapted for treating autologous
29 biological tissue for modifying its tissue reactivity and for rendering the tissue

1 temporarily more rigid than in its natural state, the composition comprising:
2 approximately 15 to 60 % by volume of ethyl alcohol;
3 approximately 2 to 10 % by weight of polyethylene glycol of a
4 molecular weight of approximately 8,000 D;
5 approximately 0.1 to 0.7 % by weight of heparine, and
6 the balance of the composition substantially consisting of water.

7 11. The liquid composition of Claim 10 comprising approximately
8 50 % ethyl alcohol, approximately 5 % polyethylene glycol and approximately
9 0.5 % heparine.

10 12. A method for modifying the tissue reactivity of an autologous
11 tissue freshly obtained from a host mammal and for rendering the tissue
12 temporarily more rigid than in its native state, the method comprising:
13 exposing said autologous tissue to a liquid composition, comprising:
14 approximately 10 to 70 % by volume of a water miscible non-toxic
15 organic solvent selected from the group consisting of an alcohol having 1 to 3
16 carbons, acetone, acetonitrile and methyl ethyl ketone;
17 approximately 2 to 30 % by weight of polyethylene glycol having a
18 molecular weight in the range of approximately 6,000 to 15,000 D;
19 approximately 0.01 to 1.0 % by weight of heparin, and
20 the balance of the composition substantially consisting of water.

21 13. The method of Claim 12 where in the liquid composition the
22 water miscible organic solvent is ethyl alcohol.

23 14. The method of Claim 13 where the liquid composition contains
24 approximately 15 to 60 % ethyl alcohol.

25 15. The method of Claim 13 where the liquid composition contains
26 approximately 2 to 10 % polyethylene glycol.

27 16. The method of Claim 13 where the liquid composition contains
28 approximately 0.1 to 0.7 % heparin.

29 17. The method of Claim 13 where the liquid composition contains

1 approximately 50 % ethyl alcohol, approximately 5 % polyethylene glycol of a
2 molecular weight of approximately 8,000 D and approximately 0.5 % heparin.

3 **18.** The method of Claim 12 where the autologous tissue freshly
4 obtained from a host mammal comprises a biological membrane.

5 **19.** The method of Claim 12 where the autologous tissue is selected
6 from a group consisting of peritoneum, pericardium, pleura and tendon.

7 **20.** The method of Claim 12 wherein the step of exposing the
8 autologous tissue to the liquid composition is by immersing the tissue in the
9 liquid composition.

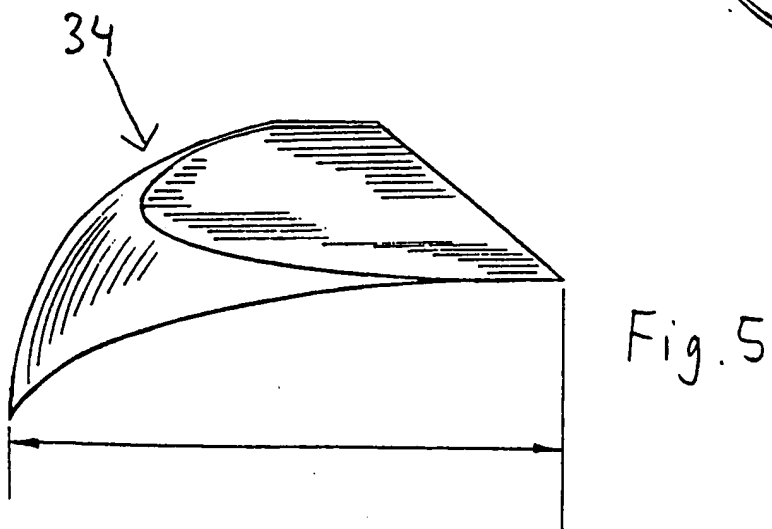
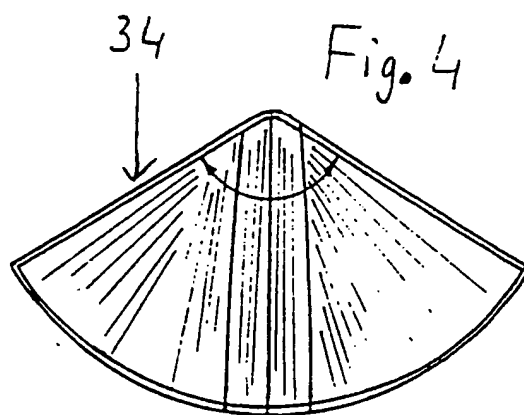
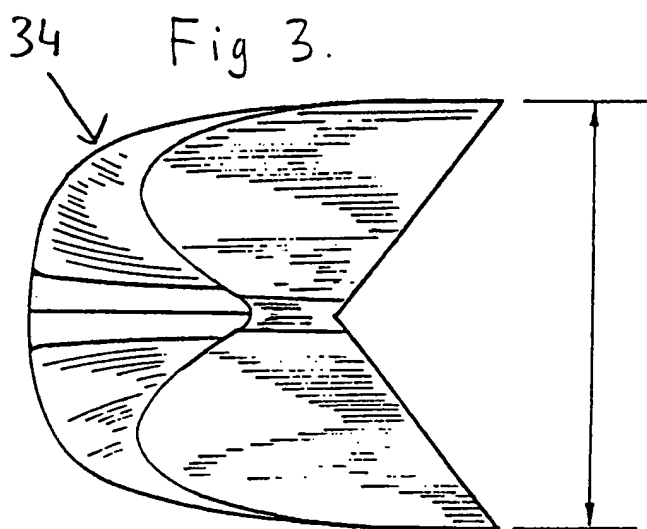
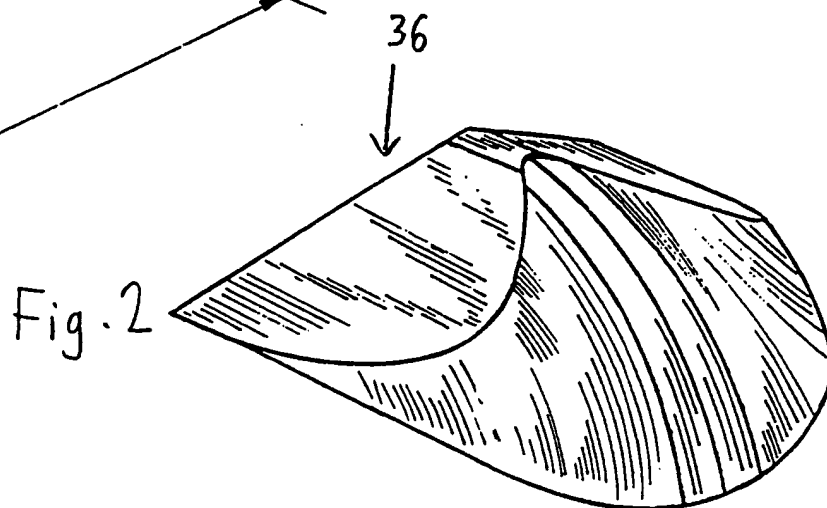
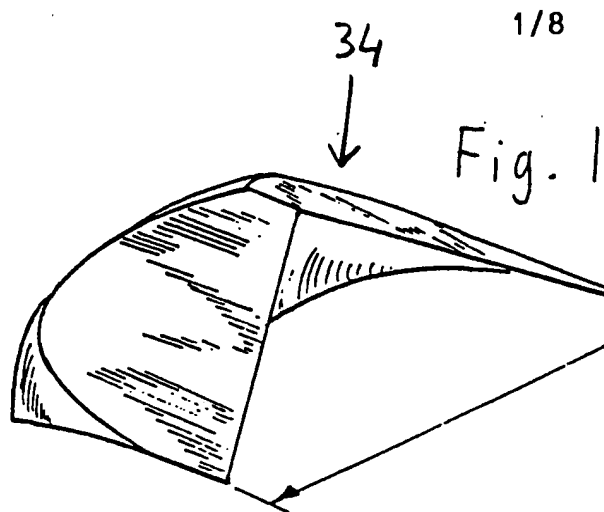
10 **21.** The method of Claim 20 further comprising the step of placing
11 the autologous tissue in a mold.

12 **22.** The method of Claim 21 wherein the autologous tissue is placed
13 in the mold before the tissue is exposed to the liquid composition, and wherein
14 the tissue is exposed to the liquid composition while it is held in said mold,
15 thereby forming said tissue into a predetermined configuration.

16 **23.** The method of Claim 22 further comprising the step of trimming
17 excess autologous tissue by cutting while said tissue is still in the mold and
18 after it has been exposed to said liquid composition for at least approximately
19 2 to 8 minutes.

20 **24.** The method of Claim 23 further comprising the step of
21 implanting the trimmed tissue into the host.

22 **25.** The method of Claim 24 further comprising the step of irrigating
23 the implanted tissue with isotonic saline solution thereby causing the physical
24 properties of the tissue to return to their substantially native state.



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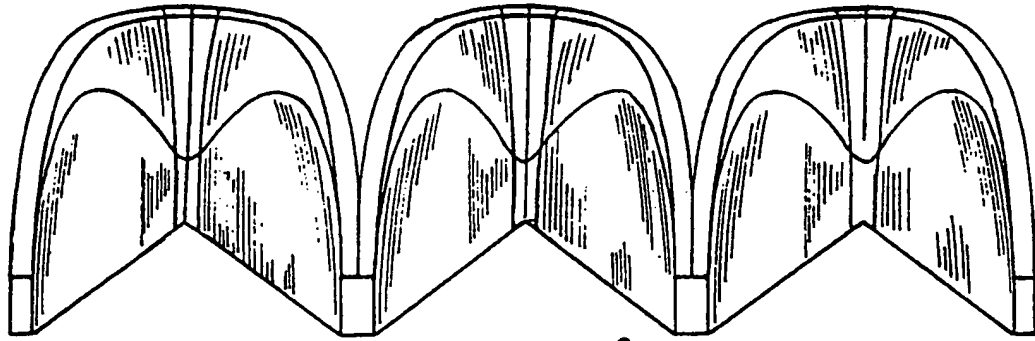
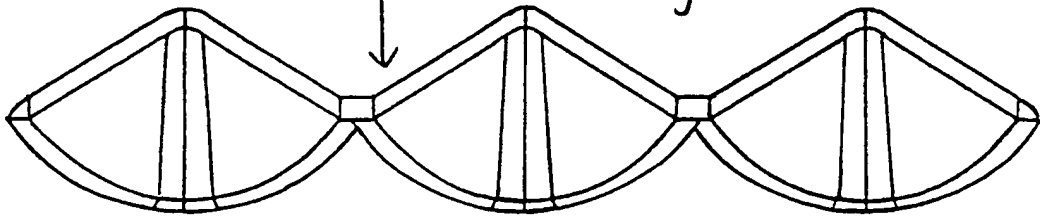


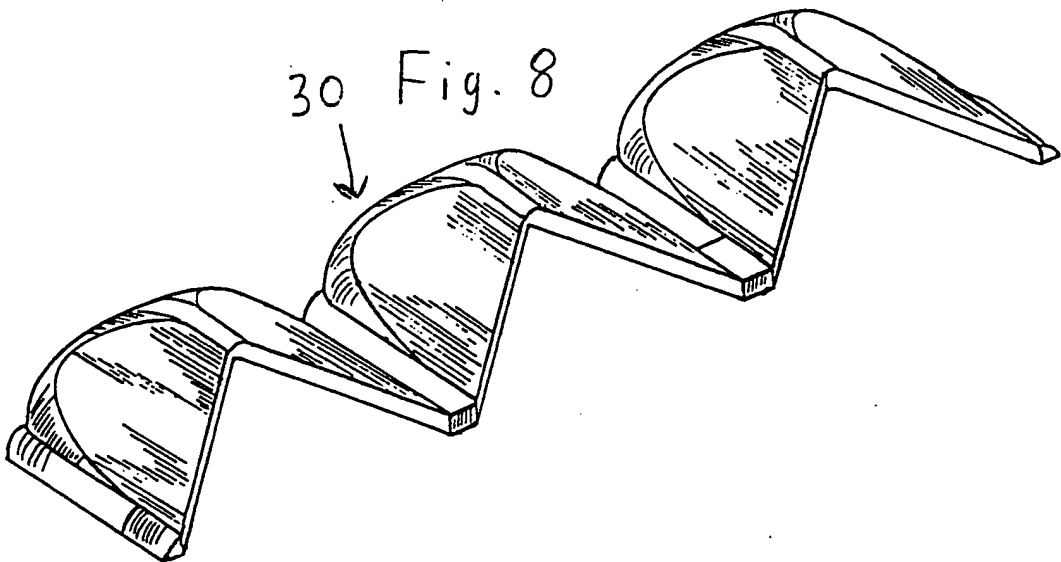
Fig. 6

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Fig 7.



30 Fig. 8



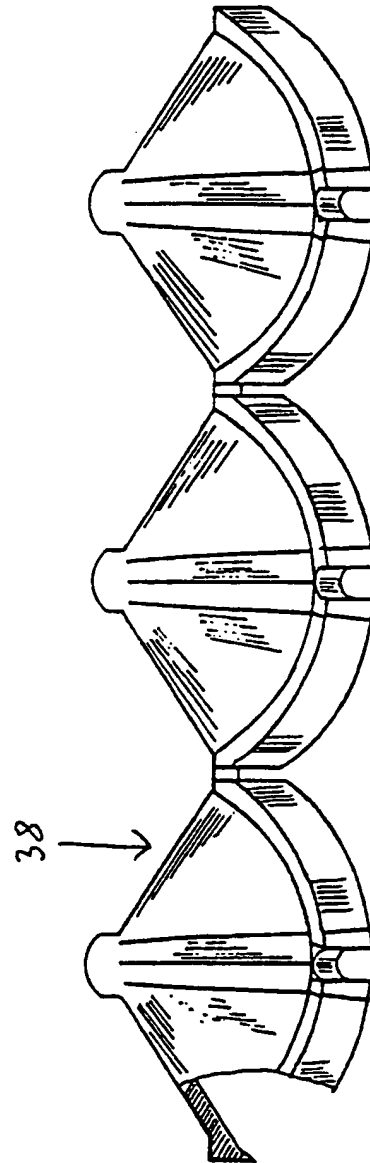
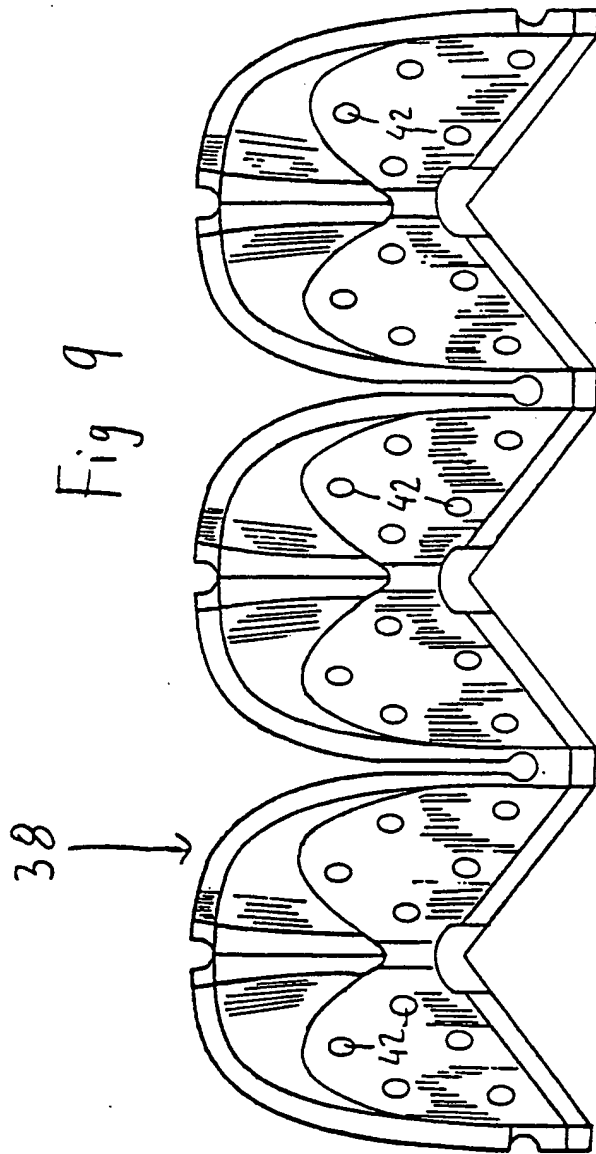


Fig. 11

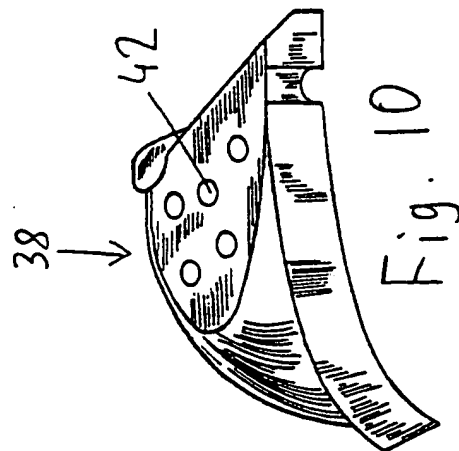


Fig. 10

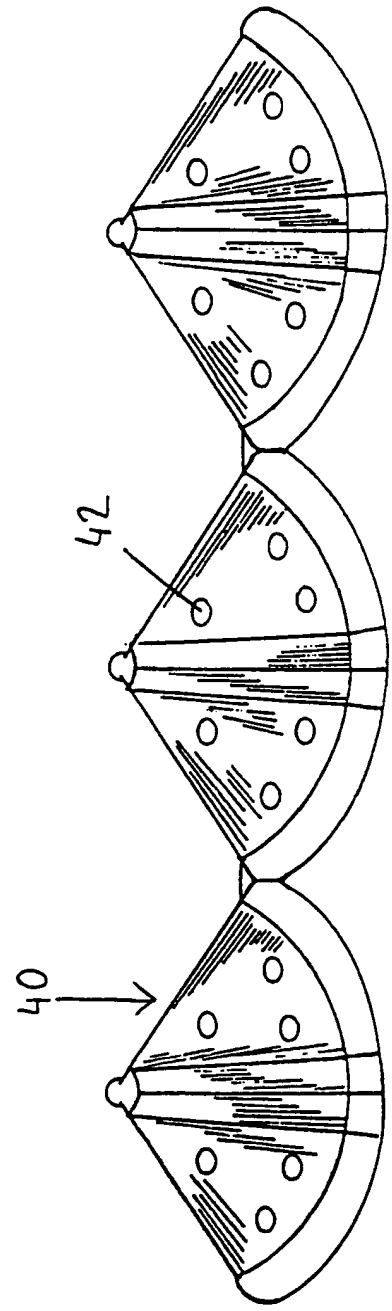
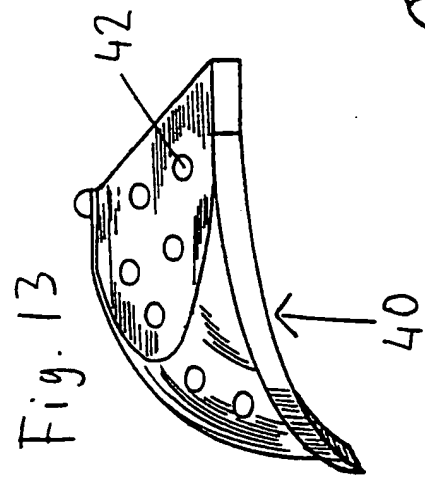
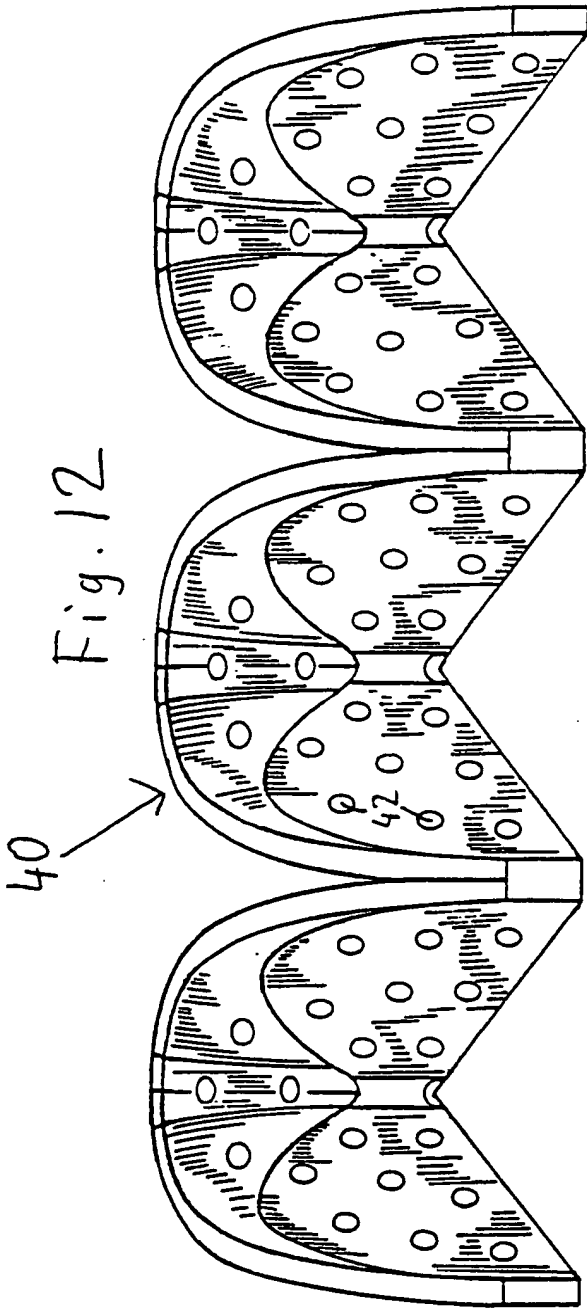
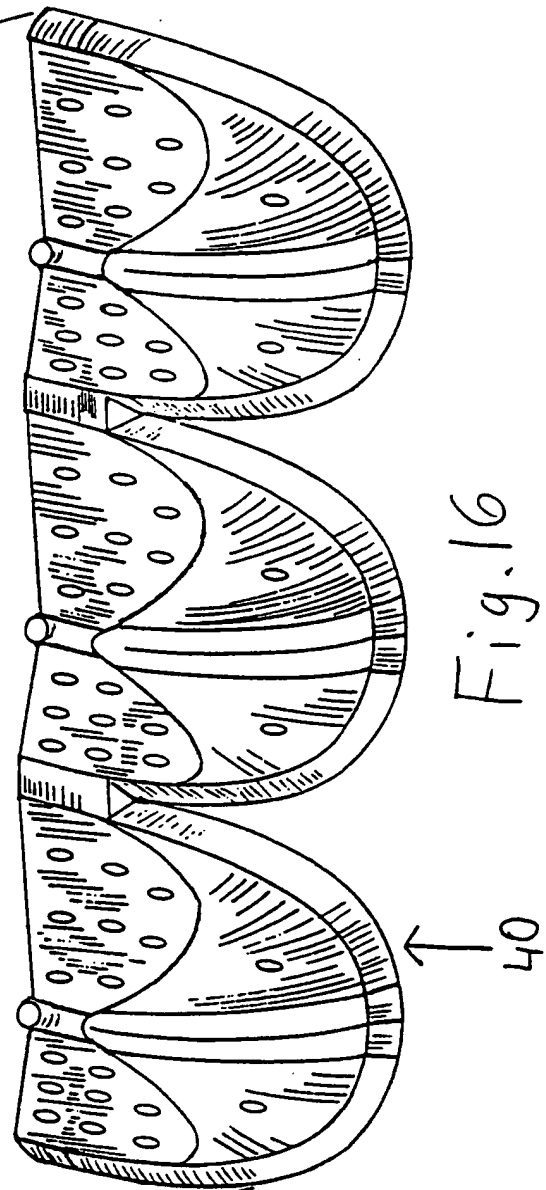
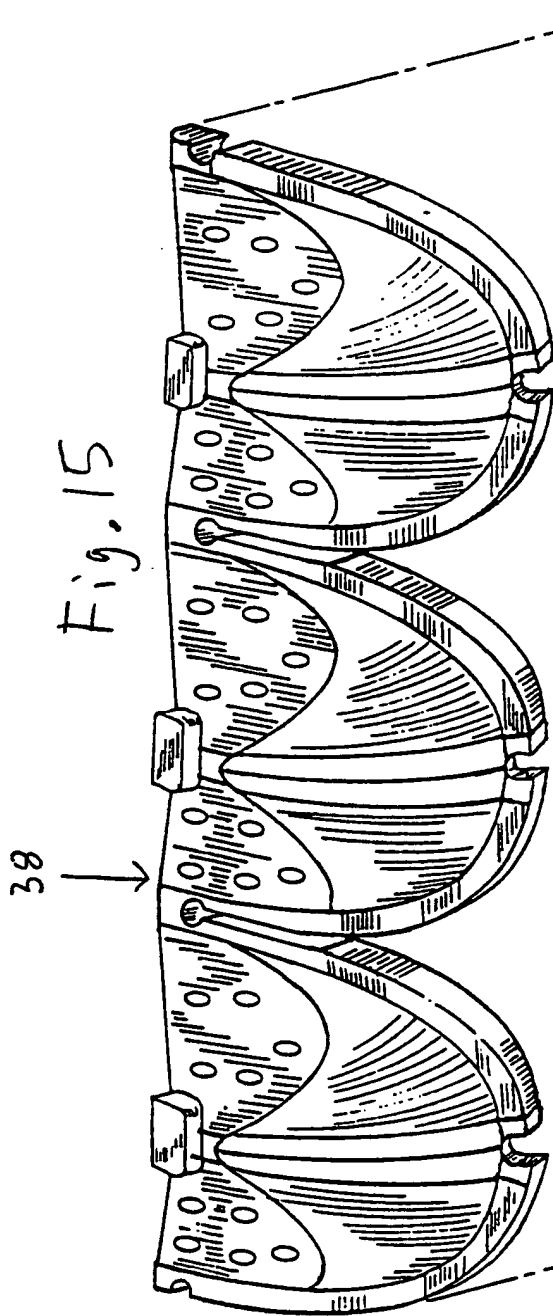
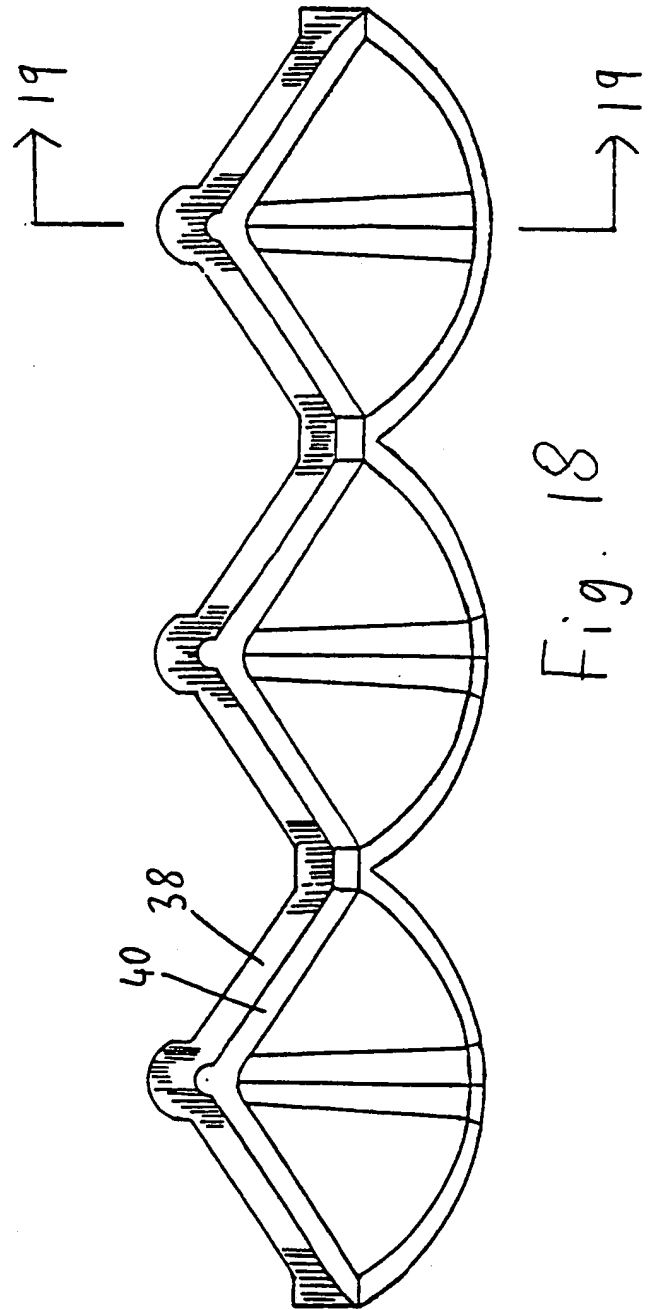
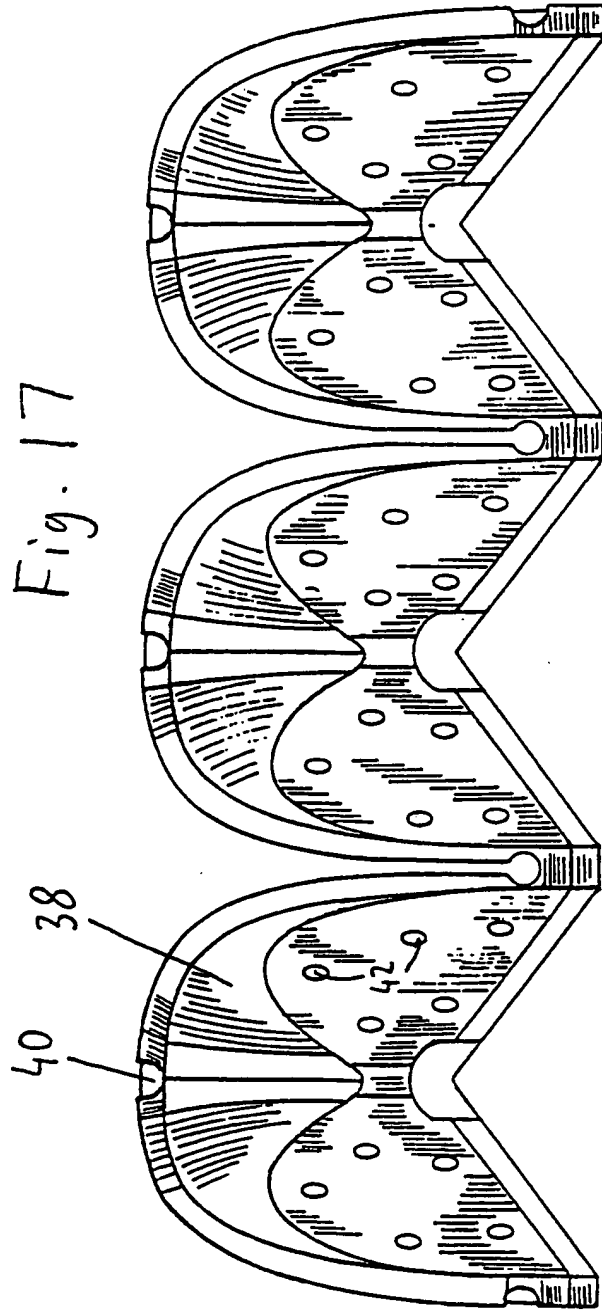


Fig. 14





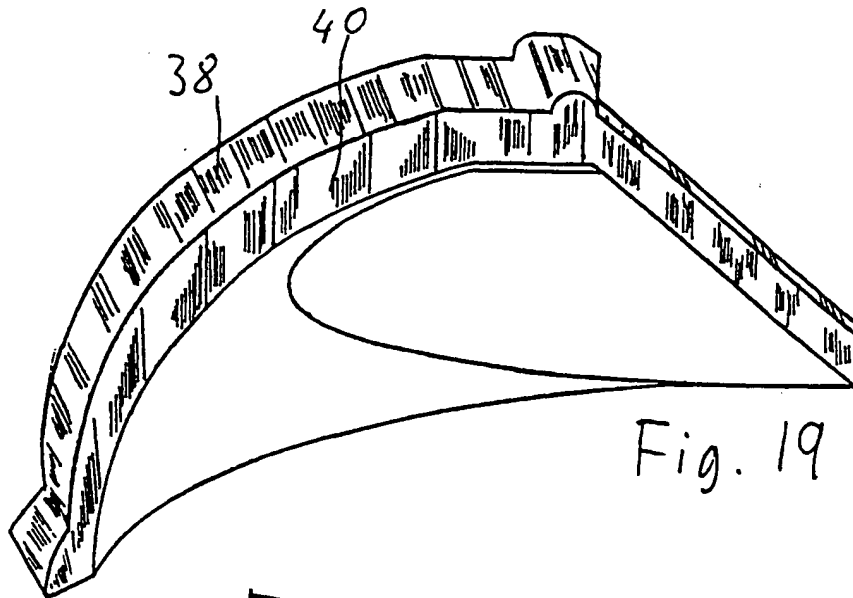


Fig. 19

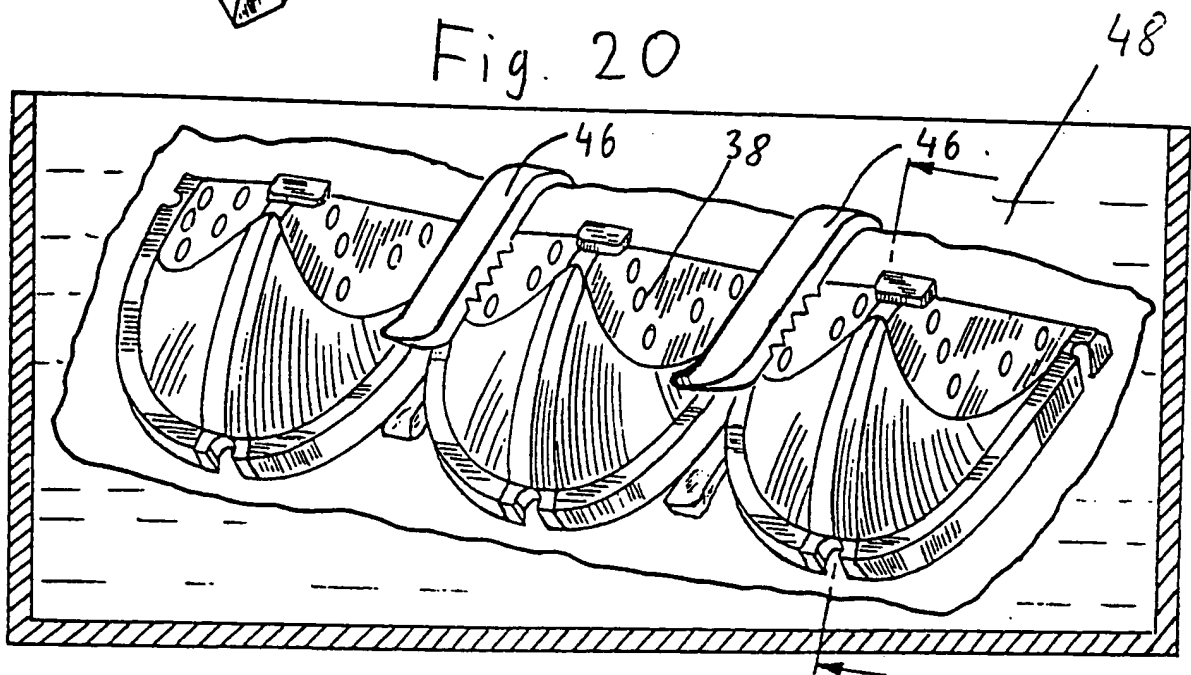


Fig. 20

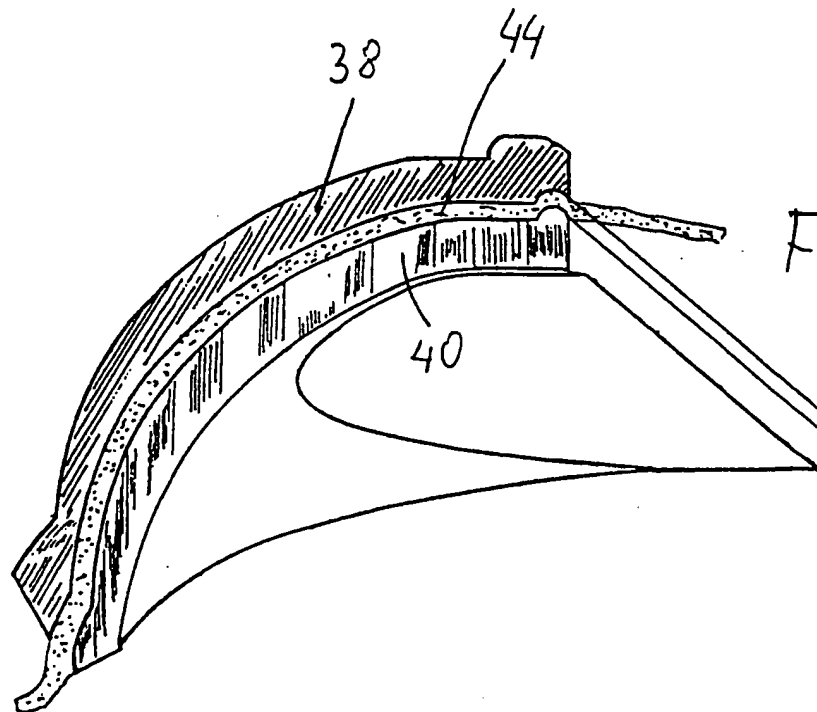
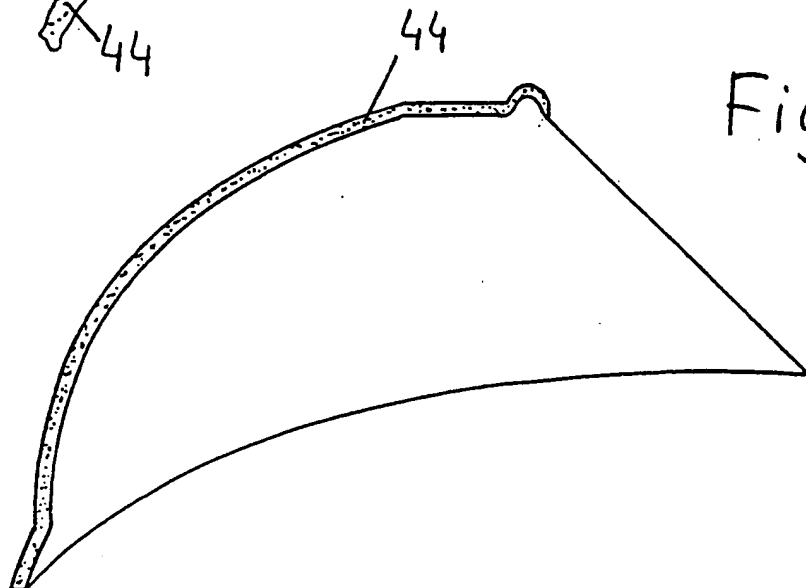
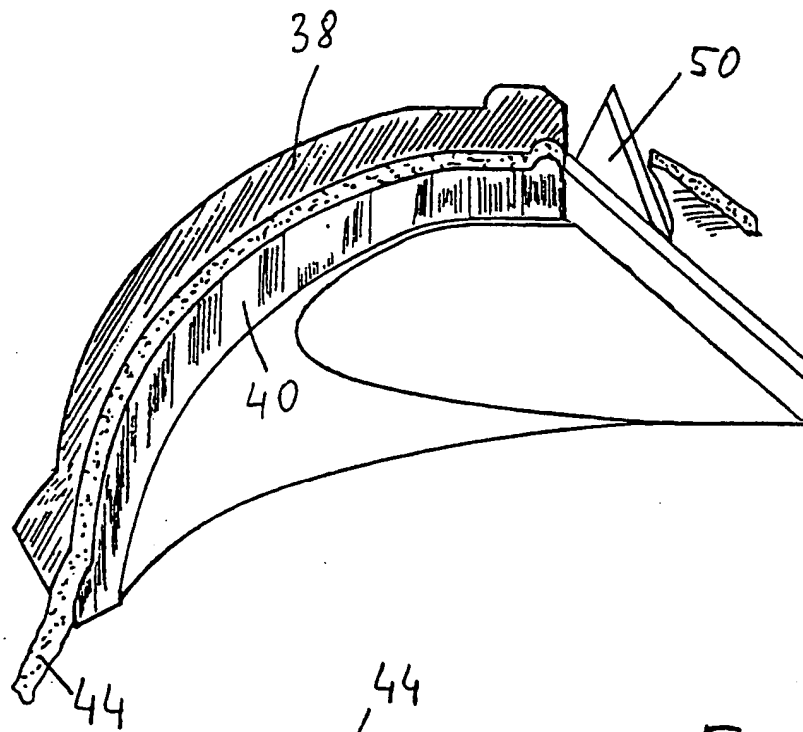
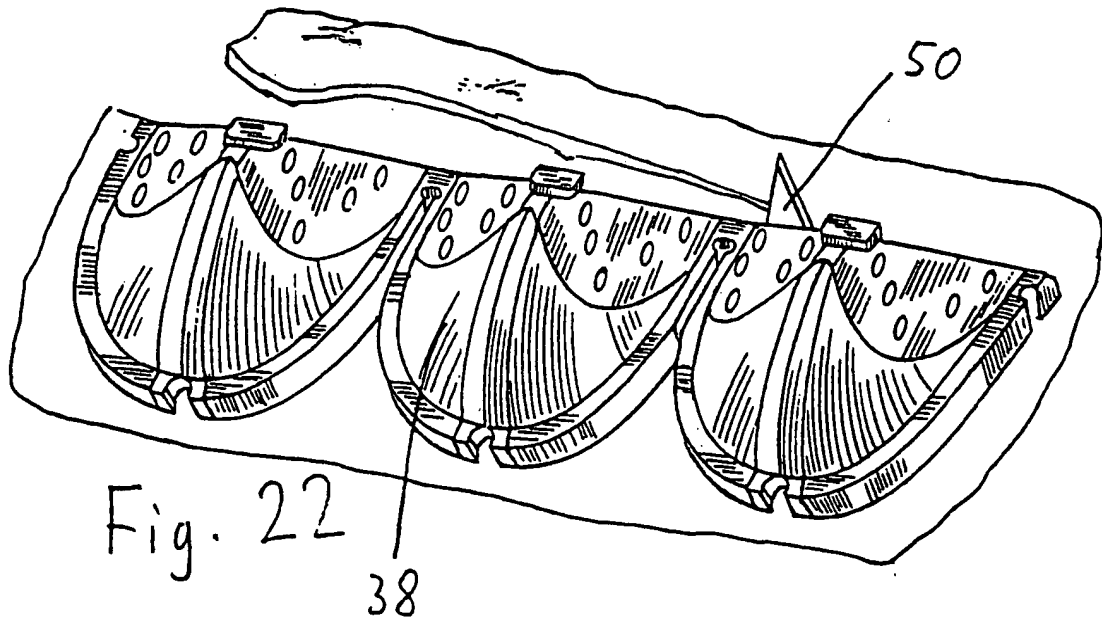


Fig. 21



INTERNATIONAL SEARCH REPORT

Int. national Application No

PCT/US 00/41142

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 A61L27/36 A01N1/02 A61F2/24

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61L A61F A01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	WO 99 66967 A (INT HEART INST OF MONTANA FOUN) 29 December 1999 (1999-12-29) the whole document ----	1-25
A	WO 98 07452 A (SULZER VASCUTEK LTD ;WALKER DONALD FRANCIS (GB)) 26 February 1998 (1998-02-26) claims; examples ----	1-25
A	US 5 558 875 A (WANG SU) 24 September 1996 (1996-09-24) cited in the application claims ----	1-25
A	WO 97 32472 A (BARBARASH LEONID SEMENOVICH ;ZHURAVLEVA IRINA JURIEVNA (RU); GANTI) 12 September 1997 (1997-09-12) abstract -----	1-25



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Patent family members are listed in annex.

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Date of the actual completion of the international search

1 March 2001

Date of mailing of the international search report

08/03/2001

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INTERNATIONAL SEARCH REPORT

Information on patent family members

I. national Application No

PCT/US 00/41142

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